



肠道菌群——运动干预治疗2型糖尿病的新靶点

赵翠翠^{1,2},覃飞^{2,3},耿雪^{2,4},徐旻霄^{2,5},瞿超艺^{2,4},赵丽娜^{2,5},赵杰修^{2*}

摘要: 肠道菌群是人体最大的微生态系统,其结构的紊乱是2型糖尿病发生发展的重要原因之一。运动作为一种内稳态刺激因素,可以通过改变宿主肠道菌群的结构多样性,影响物质能量代谢、免疫系统和神经系统等功能,从而达到防治2型糖尿病的作用。因此本文从肠道菌群的角度出发,对运动改善2型糖尿病的实验性研究及潜在机制进行总结归纳,为基于运动干预调控肠道菌群治疗2型糖尿病提供理论依据和参考。

关键词: 肠道菌群;运动;2型糖尿病

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Intestinal Flora: A New Target in the Treatment of Type 2 Diabetes by Exercise Intervention

ZHAO Cuicui^{1,2}, QIN Fei^{2,3}, GENG Xue^{2,4}, XU Minxiao^{2,5}, QU Chaoyi^{2,4}, ZHAO Lina^{2,5}, ZHAO Jiexiu^{2*}

(1. School of Sports Science, Qufu Normal University, Qufu 273165, China; 2. China Institute of Sport Science, Beijing 100061 China; 3. School of Sport, Jinan University, Guangzhou 510032, China; 4. Beijing Sport University, Beijing 100084, China; 5. Shanghai University of Sport, Shanghai 200438, China)

Abstract: Intestinal flora is the largest microecosystem in human body. Its structural disorder is one of the important causes for the occurrence and development of type 2 diabetes. As a homeostasis stimulating factor, exercise can affect the function of material and energy metabolism, immune system and nervous system by changing the structural diversity of the host intestinal flora. In this way, it can prevent and cure type 2 diabetes. Therefore, from the perspective of intestinal flora, this paper summarizes the experimental studies and potential mechanisms of exercise in improving type 2 diabetes. This provides theoretical basis and reference for the treatment of type 2 diabetes through exercise-based intervention and control of intestinal flora.

Key Words: Intestinal flora; exercise; type 2 diabetes

随着经济的快速发展,人民生活水平显著提高,久坐不动、高糖高脂的生活方式使得糖尿病等慢性代谢疾病发病率不断攀升。2014年,研究显示我国约有1.14亿糖尿病患者,2017年全球糖尿病患者的总数飙升到了4.25亿^[1,2]。糖尿病已逐渐成为世界范围内一个日益突出的社会健康问题。肠道菌群在宿主免疫系统、能量代谢、炎症以及相关基因表达等方面都至关重要。肠道微生态的平衡不仅可以促进机体的新陈代谢,还可以增强机体的免疫功能。越来越多的证据表明,肠道微生物群与糖尿病的发展密切

相关^[3,4]。肠道菌群紊乱——有益菌群的减少或有害菌群和条件致病性菌群的增强,能够使得宿主胰岛细胞功能受损、胰岛素敏感性降低、胰岛素抵抗(Insulin Resistance, IR),最终发展成2型糖尿病^[5]。

缺乏运动、肥胖以及压力等因素是导致2型糖尿病患者数量增加的主要原因。大约有27%的糖尿病、30%的缺血性心脏病、21%~25%的乳腺癌和结肠癌,可归因于缺乏体育活动。相关治疗指南建议,改变饮食和体育活动方式应作为2型糖尿病治疗的一部分^[1]。运动作为防治糖尿病最具成本效益的干预

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第一作者简介:赵翠翠,女,在读硕士研究生。主要研究方向:运动与特殊环境。E-mail:isjl228@163.com。

*通信作者简介:赵杰修,男,博士,研究员,博士生导师。主要研究方向:运动与特殊环境。E-mail:zhaojiexiu@ciss.cn。

作者单位:1.曲阜师范大学体育学院,山东曲阜273165;2.国家体育总局体育科学研究所,北京100061;3.暨南大学体育学院,广东广州510032;4.北京体育大学,北京100084;5.上海体育学院,上海200438。



方式,可在产生胰岛素抵抗的骨骼肌中诱导其保留葡萄糖摄取相关信号分子,并有效调节肠道菌群的组成和结构,提高肠道菌群的多样性和稳定性,在治疗慢性代谢性疾病中有巨大的潜力^[6]。因此,本文对运动调控肠道菌群防治2型糖尿病的相关性研究进行阐述,为基于肠道菌群防治2型糖尿病的相关研究提供思路和方法。

1 肠道菌群与2型糖尿病

正常人体内的肠道菌群主要由拟杆菌门、厚壁菌门、变形菌门、放线菌等组成^[7]。根据它们的功能,可以大致分为3类:有益菌、有害菌和条件致病菌^[8]。越来越多的证据表明,生态失调的肠道菌群在2型糖尿病的发病过程中扮演一个重要的角色^[9]。Larsen等^[5]的研究首次揭示了糖尿病患者与正常人群肠道菌群组成的显著性差异。Zhang等^[10]研究发现肠道菌群结构的变化可以调节糖尿病的发生。此外,与正常人相比,糖尿病患者肠道菌群中双歧杆菌、梭状芽孢杆菌和厚壁菌门的数量显著降低^[5,11]。在这些患者中,肠道菌群的细胞膜更活跃于糖和支链氨基酸的运输,与促炎反应直接相关的氧化应激反应蓬勃发展,从而使得宿主肠道菌群的多样性和稳定性被破坏^[12]。

目前的研究表明,肠道菌群主要通过肠道通透性增加和慢性低度炎症反应,改变短链脂肪酸(Short-Chain Fatty Acid, SCFAs)和胆汁酸(Bile Acids, BAs)的代谢扰动^[5,13]等多方面参与2型糖尿病的发生发展^[14]。一方面,肠道中不能完全被人体消化吸收的复杂碳水化合物被肠道菌群酵解,为宿主和肠道内其他共生菌的生理活动供能,同时产生SCFAs等有益代谢产物^[15]。而2型糖尿病患者体内SCFAs的减少,能影响肠道抗炎反应能力,削弱SCFAs受体激活能力,继而使宿主胰岛细胞功能受损,胰岛素敏感性随之降低,产生胰岛素抵抗(Insulin Resistance, IR),最终发展成为2型糖尿病^[16-17]。另一方面,BAs作为胆汁的组成部分,肝脏胆固醇降解产生的BAs可作为一种信号分子,在调节能量代谢、抑制肠道细菌的过度增殖等方面发挥重要作用。肠道菌群的次级BAs代谢,能够调控宿主的BAs池及代谢稳态。肠道菌群的结构异常能引发慢性低度炎症,抑制肠屏障功能,使宿主全身暴露脂多糖(Lipopolysaccharide, LPS)^[18]。LPS与单核巨噬细胞表面Toll样受体4(Toll-Like Receptor 4, TLR-4)结合,能够激活TLRs和丝裂原活化蛋白激酶(Mitogen-Activated Protein Kinase, MAPK)信号通路,促进广泛低度的慢性炎症,使得宿主发生IR,产生代谢紊乱,最终导致2型糖尿病^[19]。

2 运动对2型糖尿病患者肠道菌群的影响

运动作为一种内稳态刺激,可能使肠道菌群多样化,增加良性微生物群落的数量^[20-21]。运动时肠道菌群定性和定量的改变能够影响营养物质的吸收、能量分布、免疫和内脏—大脑轴^[22]。多项研究表明,即使中等强度的体育活动也可改善机体新陈代谢和整体健康。运动通过改变肠道菌群的组成和肠道屏障功能,控制糖尿病的发生发展^[23]。然而不同强度的体育活动会对机体肠道菌群造成不同的影响。运动与微生物群之间的关系是复杂的,它取决于运动的强度以及时间等多项因素,已经观察到不同运动强度对2型糖尿病患者有益和有害的影响。

2.1 中小强度运动

多项研究结果显示中小强度运动对机体存在有益的效果。长期(12个月)中强度的有氧运动和抗阻运动能够减少白细胞介素-1 β (Interleukin-1 β , IL-1 β)、肿瘤坏死因子- α (Tumor Necrosis Factor- α , TNF- α)等炎性细胞因子,增加IL-4等抗炎性细胞因子^[24]。增加中等强度的习惯性体力活动时间对GLP-1的分泌有好处,这有助于改善对葡萄糖的调节,降低2型糖尿病的发生风险^[25]。Lambert等^[26]的一项动物模型实验数据表明,糖尿病小鼠进行6周的运动后,小鼠肠道菌群的组成发生了有利变化,包括拟杆菌门的减少,厚壁菌门和变形杆菌门的增加^[27],进而使SCFAs和BAs含量增加^[22]。在2型糖尿病患者中,与能量消耗匹配的持续步行训练相比,间隔步行训练有助于维持胰岛素分泌,改善胰岛素敏感性^[28]。一项对30例临床稳定型2型糖尿病患者进行为期6个月的耐力、阻力和柔韧性训练的研究结果显示,慢性运动能够改善由糖尿病引起的肠道菌群过度生长、肠道通透性增加和全身低度炎症。这些密切相关的变量表明运动还可以通过改变肠道菌群组成和肠道屏障功能来调控糖尿病。此研究也显示了慢性运动的一个额外机制,提示改善肠道菌群可能是2型糖尿病个体化治疗的重要一步^[29]。太极作为一种包括深呼吸和冥想的中等强度的运动,也可能通过调节下丘脑—垂体—肾上腺(Hypothalamic-Pituitary-Adrenal, HPA)而引起肠道菌群的有益变化^[30]。在有氧运动后,正常体重患者的厌食代谢会升高,食欲(胃饥饿素和瘦素)降低^[31]。有氧运动引起的线粒体生物发生,以转录共激活因子如PCG-1的细胞活化和氧化还原敏感能量传感器(SIRT1)为特征,也调节肠屏障功能和黏膜免疫应答^[32-33]。然而强迫游泳导致氨基丁酸显著降低,HPA和代谢过程被激活,增加肠上



皮通透性和肠炎症反应^[34],对机体造成不良影响。需要注意的是,有氧运动对肠道菌群的好处似乎是独立于饮食的,并在重新开始一种久坐的生活方式后消失。

2.2 大强度运动

相关人体实验研究也表明,职业运动员的微生物群落具有更高的多样性和更良好的代谢能力^[35-36]。经常进行高强度运动的人体内拥有健康的微生物群,可以改善代谢性疾病^[36]。在高强度运动的情况下,肠道黏膜的缺血效应以及细菌和毒素的通透性增加可导致炎症反应,这在常规运动后是不存在的^[37]。肠道屏障是由肠细胞膜、紧密连接蛋白和免疫因子(巨噬细胞)构成的^[22]。当体育锻炼超过70%VO_{2max}时,HPA的激活加上对肠黏膜的直接热损伤,导致肠血流量减少,增加对内毒素的通透性^[31]。在剧烈的运动中,血液从胃肠道流经周围器官(肌肉、心脏和肺)会导致紧密连接蛋白的松动,从而使得肠道屏障受损^[22]。在剧烈的体育活动中,缺血导致的肠道高渗透会诱导炎症级联反应(TNF和IL-6增加),从而抑制肠道AMPK的激活^[38],该代谢途径的抑制可降低葡萄糖代谢、游离脂肪酸氧化,并增加线粒体中活性氧的生成^[39-40]。有证据表明,在长时间的高强度运动训练中,相关的免疫反应会被抑制^[41]。与低强度运动相比,剧烈运动导致促炎细胞因子数量的增加,TNF- α 、IL-1等^[36,42]。众所周知,剧烈运动会增加运动员上呼吸道感染和消化系统疾病的发病率^[43]。然而两项侧重于对运动员高强度训练的研究调查了半程马拉松对其的影响,刚跑完半程马拉松后,微生物群落的多样性并没有增加,而在马拉松结束后的几天,微生物群发生了有益的变化,其中韦荣氏球菌属最为丰富^[44-45]。

短时间(5 min)中等强度的运动可能有益于肠道菌群。相反,长距离耐力运动可能通过减少肠血流量引起黏膜糜烂和缺血性结肠炎^[46]。多项研究引出了一个理论,即存在免疫受损的“打开的窗口”^[47],在这个窗口中,病毒和细菌更有可能增加耐力运动员亚临床和临床感染的风险。运动的时间和强度可能有一个阈值,通过该阈值可以改善结肠功能,这需要在后续的研究中进一步确认。因此,相关学者建议2型糖尿病患者应进行每周150 min的中至高强度运动或每周75 min的高强度运动或间隔训练,至少每周3 d,以降低2型糖尿病相关并发症的发生风险^[48]。总之,生活方式的改变,尤其是运动对血糖控制和预防糖尿病相关并发症的重要性不言而喻。

3 运动改善肠道菌群防治2型糖尿病的潜在机制

3.1 对宿主能量代谢的影响

越来越多的研究显示,肠道菌群的能量代谢异常是2型糖尿病发生发展的重要原因之一,肠道乳酸菌和双歧杆菌等益生菌糖耐量紊乱,影响葡萄糖的吸收,同时促进脂肪的合成和储存,并参与糖尿病的发生^[14]。然而,肠道菌群的有益代谢产物SCFAs、BAs等在宿主能量代谢中发挥重要作用^[14]。

3.1.1 SCFAs

SCFAs是人体大肠上皮细胞主要的能量来源^[49],主要包括乙酸盐、丙酸盐、丁酸盐等。丁酸盐为结肠上皮细胞提供能量,乙酸盐和丙酸盐到达肝脏和周围器官,是糖异生和脂肪生成的底物^[50]。

SCFAs通过调节免疫系统,减少炎症和氧化应激,促进肠道能量平衡在宿主的代谢和免疫中发挥重要作用^[51]。SCFAs可激活G蛋白偶联受体41(G-protein-coupled receptor 41,GPR41),促进肠上皮淋巴细胞(Intestinal Intraepithelial Lymphocytes,IELs)分泌多肽YY激素(Peptide YY,PYY),参与食物的摄取;也能够激活GPR43,调节IELs胞浆内Ca²⁺平衡,促进GLP-1的分泌。因此低水平的SCFAs可使机体PYY分泌减少,导致食物摄取过量,胰岛素分泌减少,胰岛素敏感性降低^[52-53],甚至产生IR^[54]。Estaki等^[55]研究表明,心肺适能与微生物多样性的增加和丁酸盐的产生有关。有关太极拳与肠道菌群的一项研究表明,太极拳可以通过改善心肺健康来调节肠道微生物群^[56]。相关实验研究表明,体育活动能够改变粪便短链脂肪酸(SCFAs),增加粪便丁酸盐的存在,进而增加产生丁酸盐的肠道细菌^[57]。SCFAs(尤其是丙酸盐)可以诱导PYY和GLP-1的分泌,这两种肠内分泌激素对能量平衡和葡萄糖稳态至关重要^[58]。GLP-1能够增强高血糖状态下的胰岛素分泌,并抑制胃排空、食物摄入和胰高血糖素分泌,从而降低2型糖尿病患者的血糖水平^[59]。急性高强度运动会导致短暂的食欲下降,抑制胃促生长素,增加GLP-1的浓度^[60]。有趣的是,SCFAs能够激活肌肉AMPK,调节肌肉葡萄糖和脂质代谢,减轻脂肪堆积^[61]。Islam等^[60]证明,健康被试的剧烈运动通过乳酸和IL-6调节食欲,导致GLP-1分泌,从而达到能量平衡。这些代谢效应在糖尿病的发展中至关重要,并证实了微生物群、运动和整体代谢之间的关系。



3.1.2 BAs

BAs作为一种信号分子,不仅在脂肪和脂溶性维生素的吸收、转运和分布中起重要作用,还能够调节能量代谢,从而抑制肠道菌群的过度增殖。菌群失调导致BAs活化减少,从而降低游离的BAs和次级BAs水平,法尼醇受体(Farnesoid X Receptor, FXR)和G蛋白偶联胆汁酸受体5(G Protein-Coupled Bile Acid Receptor 5, TGR5)的活化减弱,可抑制糖异生基因的表达,进而减少胰岛素的分泌、降低胰岛素的敏感性、食欲增加和肥胖,最终导致2型糖尿病的发生。相关研究表明,肠道拟杆菌以及大肠杆菌参与了结肠中次级BAs的生成^[62],那么运动可能通过增加有益菌群的含量,来增加结肠中次级BAs的含量。运动干预通过降低TLR4依赖性炎症反应,下调核因子激活的B细胞的K-轻链增强的转录活性来保持正常肠道屏障功能,反过来阻止了肝-肠的失调,进而改善了BAs稳态^[63]。Meissner等^[64]发现,与久坐不动的小鼠相比,使用滚轮运行12周的小鼠的BAs分泌增加,粪便BAs输出增加。粪便BAs随着运动量和强度的增加而增加^[65]。然而运动的方式、强度的阈值有待进一步研究。

3.2 对宿主免疫系统的影响

研究表明,肠道淋巴细胞的抗炎状态、调节肠道屏障功能障碍、保留黏膜厚度和肠道通透性与有规律的身体活动相关^[66]。肠道菌群紊乱能够导致内分泌调节肽分泌减少,肠上皮淋巴细胞IELs活化增加,最终诱发2型糖尿病。

3.2.1 炎症方面

TLR-4能够识别脂多糖与葡萄糖磷酸异构酶锚定蛋白CD14形成的复合物,进而作用于骨骼肌细胞和脂肪细胞,通过TLRs和MAPK信号通路促进炎症和IR的发展^[14]。

运动介导的器官串扰形成一种模式,导致抗炎细胞因子的增加和/或促炎细胞因子的减少。一方面运动刺激肌肉肌素的释放,通过激活AMPK相关信号通路,降低血清脂多糖(Lipopolysaccharide, LPS)水平来抑制肝脏、肌肉和脂肪组织中的TLRs信号通路来增加肌肉葡萄糖代谢进而降低机体炎症^[67-68]。特别是急性运动产生的IL-6被认为是调节免疫和炎症代谢的多效性肌动蛋白:它抑制TNF- α 产生^[69],同时刺激抗炎细胞因子IL-1 α 和IL-10的释放^[70-71]。另一方面,运动能够增加肠道有益菌群的含量进而提高丁酸盐等SCFAs的含量,通过调节小鼠T细胞的分化^[72]、增加免疫球蛋白A的产生^[73],抑制

促炎因子的产生,促进黏膜炎症的修复,进而抑制结肠的炎症反应;SCFAs通过调节中性粒细胞来抑制炎症反应^[15,49],促进调控食欲相关中枢肽以及肠促胰岛素的分泌^[51-52,74,75],从各种神经内分泌及表观遗传机制方面对人的食欲进行调控并影响胃肠排空。

3.2.2 肠屏障方面

高脂饮食、久坐不动等因素使得乳酸杆菌等有益菌群比例下降,革兰氏阴性细菌的比例上升,肠道菌群的紊乱导致肠壁通透性上升。代谢内毒素血症是通过产生和吸收更多的脂多糖,抑制肠道物理屏障、微生物屏障功能而产生的^[14]。

与久坐不动的人相比,训练有素的运动员在休息时细菌内毒素脂多糖水平更低^[76],对热应激的热休克蛋白反应更强^[77]。肠道中增加的热休克蛋白已被证明可以防止上皮细胞间紧密连接蛋白的破坏^[78]。减少细菌移位和提高抗微生物蛋白的产生也与适度运动有关。SCFAs可以改善结肠的酸性环境,抑制有害细菌的生长,维持水和电解质的平衡,诱导结肠黏膜通透性降低^[59]。因此,运动作为一种有益的刺激,能够提高肠道屏障的长期弹性,从而改善2型糖尿病^[79]。

4 总结与展望

多项研究证明肠道菌群与宿主免疫、能量代谢等相关的生理活动息息相关,肠道微生物的失衡与2型糖尿病等多种代谢疾病的发生发展密不可分。肠道生态系统的紊乱可导致有益菌丰度降低,有害菌比例增加,从而导致SCFAs、BAs与内分泌调节肽的含量降低,影响机体肠道氧化还原稳态以及肠黏膜的屏障功能。多项研究表明,运动能够增加有益菌的比例,从而提高SCFAs和BAs的含量,SCFAs和BAs又能够促进相关内分泌调节肽如GLP-1的分泌。运动能够激活AMPK相关信号通路,降低机体炎症;肌源性IL-6能够抑制肿瘤坏死因子的产生,同时能够增加IL-10等抗炎因子的产生,使得肠上皮细胞的代谢稳态恢复平衡,从而提高胰岛素敏感性,最终改善2型糖尿病。

目前,虽然越来越多的证据表明,运动是预防和治疗糖尿病绿色安全、最具成本效益的生活干预手段,不会发生像粪移植后的多重耐药菌的严重感染甚至死亡的风险。体育锻炼能够通过调节肠道微生物群改善2型糖尿病,但是其中的机制远远未被清楚地阐明。许多实验研究忽略了饮食等一系列相关因素,运动的形式、强度和持续时间仍是未来研究的重点。因此,在今后的具体研究方案中应该结合患者的情况,探索最佳的运动治疗方案。



参考文献:

- [1] Cho N. H., Shaw J. E., Karuranga S., et al. IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045[J]. *Diabetes Res. Clin. Pr.*, 2018, 138(2):71-81.
- [2] Wang L. M., Gao P., Zhang M., et al. Prevalence and ethnic pattern of diabetes and prediabetes in China in 2013 [J]. *J. Am. Med. Assoc.*, 2017, 317(24): 2515-2523.
- [3] Haro C., Montes-Borrego M., Rangel-Zuniga O. A., et al. Two healthy diets modulate gut microbial community improving insulin sensitivity in a human obese population [J]. *J. Clin. Endocr. Metab.*, 2016, 101(1): 232-241.
- [4] Mejia-Leon M. E., de La Barca A. M. C. Diet, microbiota and immune system in type 1 diabetes development and evolution[J]. *Nutrients*, 2015, 7(11): 9171-9184.
- [5] Larsen N., Vogensen F. K., van Den Berg F. W., et al. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults[J]. *PLoS One*, 2010, 5(2): e9085.
- [6] Sylow L., Kleinert M., Richter E. A., et al. Exercise-stimulated glucose uptake - regulation and implications for glycaemic control[J]. *Nat Rev Endocrinol*, 2017, 13(3): 133-148.
- [7] Tremaroli V., Backhed F. Functional interactions between the gut microbiota and host metabolism[J]. *Nature*, 2012, 489(7415): 242-249.
- [8] Guarner F., Malagelada J. R. Gut flora in health and disease[J]. *Lancet*, 2003, 361(9356): 512-519.
- [9] Bouter K. E., van Raalte D. H., Groen A. K., et al. Role of the gut microbiome in the pathogenesis of obesity and obesity-related Metabolic Dysfunction [J]. *Gastroenterology*, 2017, 152(7): 1671-1678.
- [10] Zhang X. Y., Shen D. Q., Fang Z. W., et al. Human gut microbiota changes reveal the progression of glucose intolerance[J]. *PLoS One*, 2013, 8(8): e71108.
- [11] Qiu J., Zhou H., Jing Y., et al. Association between blood microbiome and type 2 diabetes mellitus: A nested case-control study [J]. *J. Clin. Lab. Anal.*, 2019, 33(4): e22842.
- [12] Pedersen H. K., Gudmundsdottir V., Nielsen H. B., et al. Human gut microbes impact host serum metabolome and insulin sensitivity [J]. *Nature*, 2016, 535(7612): 376-381.
- [13] Utschneider K. M., Kratz M., Damman C. J., et al. Mechanisms linking the gut microbiome and glucose metabolism [J]. *J. Clin. Endocr. Metab.*, 2016, 101(6): 2622.
- [14] Ma Q. T., Li Y. Q., Li P. F., et al. Research progress in the relationship between type 2 diabetes mellitus and intestinal flora [J]. *Biomed Pharmacother*, 2019, 117:109-138.
- [15] Conterno L., Fava F., Viola R., et al. Obesity and the gut microbiota: Does up-regulating colonic fermentation protect against obesity and metabolic disease? [J]. *Genes Nutr.*, 2011, 6(3): 241-260.
- [16] Karlsson F. H., Tremaroli V., Nookaew I., et al. Gut metagenome in European women with normal, impaired and diabetic glucose control [J]. *Nature*, 2013, 498(7452):99-103.
- [17] Qin J. J., Li Y. R., Cai Z. M., et al. A metagenome-wide association study of gut microbiota in type 2 diabetes [J]. *Nature*, 2012, 490(7418): 55-60.
- [18] Cani P. D., Neyrinck A. M., Fava F., et al. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia[J]. *Diabetologia*, 2007,50 (11): 2374-2383.
- [19] Cani P. D., Amar J., Iglesias M. A., et al. Metabolic endotoxemia initiates obesity and insulin resistance [J]. *Diabetes*, 2007, 56(7): 1761-1772.
- [20] Bermon S., Petriz B., Kajeniene A., et al. The microbiota: An exercise immunology perspective [J]. *Exerc. Immunol Rev.*, 2015, 21(21):70.
- [21] Mika A., van Treuren W., Gonzalez A., et al. Exercise is more effective at altering gut microbial composition and producing stable changes in lean mass in juvenile versus adult male F344 Rats [J]. *PLoS One*, 2015, 10(5): e0125889.
- [22] Codella R., Luzi L., Terruzzi I. Exercise has the guts: How physical activity may positively modulate gut microbiota in chronic and immune-based diseases [J]. *Digestive and Liver Disease*, 2018, 50(4): 331-341.
- [23] Pasini E., Corsetti G., Assanelli D., et al. Effects of chronic exercise on gut microbiota and intestinal barrier in human with type 2 diabetes [J]. *Minerva Med.*, 2019, 110(1): 3-11.
- [24] Balducci S., Zanuso S., Nicolucci A., et al. Anti-inflammatory effect of exercise training in subjects with type 2 diabetes and the metabolic syndrome is dependent on exercise modalities and independent of weight loss [J]. *Nutr. Metab. Cardiovasc. Dis.*, 2010, 20(8): 608-617.
- [25] Janus C., Vistisen D., Amadid H., et al. Habitual physical activity is associated with lower fasting and greater glucose-induced GLP-1 response in men [J]. *Endocr. Connect*, 2019, 8(12): 1607-1617.
- [26] Lambert J. E., Myslicki J. P., Bomhof M. R., et al. Exercise training modifies gut microbiota in normal and diabetic mice [J]. *Appl Physiol Nutr Me*, 2015, 40(7): 749-752.
- [27] Choi J. J., Eum S. Y., Rampersau D. E., et al. Exercise attenuates PCB-Induced changes in the mouse gut microbiome [J]. *Environ. Health Persp.*, 2013, 121(6):725-730.
- [28] Karstoft K., Clark M. A., Jakobsen I., et al. The effects



- of 2 weeks of interval vs continuous walking training on glycaemic control and whole-body oxidative stress in individuals with type 2 diabetes: A controlled, randomised, crossover trial[J]. *Diabetologia*, 2017, 60(3): 508-517.
- [29] Pasini E., Corsetti G., Assanelli D., et al. Effects of chronic exercise on gut microbiota and intestinal barrier in human with type 2 diabetes [J]. *Minerva Med.*, 2019, 110(1): 3-11.
- [30] Hamasaki H. Exercise and gut microbiota: Clinical implications for the feasibility of Tai Chi [J]. *J. Integr. Med.*, 2017, 15(4): 270-281.
- [31] Zouhal H., Sellami M., Saeidi A., et al. Effect of physical exercise and training on gastrointestinal hormones in populations with different weight statuses [J]. *Nutr. Rev.*, 2019, 77(7): 455-477.
- [32] Dalton A., Mermier C., Zuhl M. Exercise influence on the microbiome-gut-brain axis [J]. *Gut Microbes*, 2019, 10(5): 555-568.
- [33] Radak Z., Torma F., Berkes I., et al. Exercise effects on physiological function during aging [J]. *Free Radic. Biol. Med.*, 2019, 132:33-41.
- [34] Martin C. R., Osadchiy V., Kalani A., et al. The brain-gut-microbiome axis [J]. *Cellular and Molecular Gastroenterology and Hepatology*, 2018, 6(2): 133-148.
- [35] Barton W., Penney N. C., Cronin O., et al. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level [J]. *Gut*, 2018, 67(4): 625-633.
- [36] Clarke S. F., Murphy E. F., O'Sullivan O., et al. Exercise and associated dietary extremes impact on gut microbial diversity [J]. *Gut*, 2014, 63(12): 1913-1920.
- [37] Karl J. P., Margolis L. M., Madslie E. H., et al. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress [J]. *Am. J. Physiol. Gastrointest Liver Physiol.*, 2017, 312(6): 559-571.
- [38] Holland A. M., Hyatt H. W., Smuder A. J., et al. Influence of endurance exercise training on antioxidant enzymes, tight junction proteins, and inflammatory markers in the rat ileum [J]. *BMC Res. Notes*, 2015, 8(1):514.
- [39] de Zoete M. R., Flavell R. A. Interactions between nod-like receptors and intestinal bacteria [J]. *Front Immunol*, 2013, 4:462.
- [40] Saint-Georges-Chaumet Y., Attaf D., Pelletier E., et al. Targeting microbiota-mitochondria inter-talk: Microbiota control mitochondria metabolism [J]. *Cell Mol Biol (Noisy-le-grand)*, 2015, 61(4): 121-124.
- [41] Shephard R. J., Shek P. N. Potential impact of physical activity and sport on the immune system—a brief review [J]. *Br. J. Sports Med.*, 1994, 28(4): 247-255.
- [42] Pedersen B. K., Toft A. D. Effects of exercise on lymphocytes and cytokines [J]. *Br. J. Sports Med.*, 2000,34(4): 246-251.
- [43] Gleeson M. Immune function in sport and exercise [J]. *J. Appl. Physiol.* 2007, 103(2): 693-699.
- [44] Zhao X., Zhang Z., Hu B., et al. Response of gut microbiota to metabolite changes induced by endurance exercise [J]. *Front Microbiol*, 2018, 9:1-11.
- [45] Scheiman J., Luber J. M., Chavkin T. A., et al. Metagenomics analysis of elite athletes identifies a performance-enhancing microbe that functions via lactate metabolism [J]. *Nat. Med.*, 2019, 25(7): 1104-1109.
- [46] Choi S. C., Choi S. J., Kim J. A., et al. The role of gastrointestinal endoscopy in long-distance runners with gastrointestinal symptoms [J]. *Eur. J. Gastroenterol Hepatol*, 2001, 13(9): 1089-1094.
- [47] Kekkonen R. A., Vasankari T. J., Vuorimaa T., et al. The effect of probiotics on respiratory infections and gastrointestinal symptoms during training in marathon runners [J]. *Int. J. Sport Nutr. Exerc. Metab.*, 2007, 17(4): 352-363.
- [48] American Diabetes A. 4 lifestyle management: Standards of medical care in diabetes-2018 [J]. *Diabetes Care*, 2018, 41(1): 38-50.
- [49] Gao Z. G., Yin J., Zhang J., et al. Butyrate improves insulin sensitivity and increases energy expenditure in mice [J]. *Diabetes*, 2009, 58(7): 1509-1517.
- [50] Marchesi J. R., Adams D. H., Fava F., et al. The gut microbiota and host health: A new clinical frontier [J]. *Gut*, 2016, 65(2): 330-339.
- [51] Koh A., de Vadder F., Kovatcheva-Datchary P., et al. From dietary fiber to host physiology: Short-chain fatty acids as key bacterial metabolites [J]. *Cell*, 2016, 165(6): 1332-1345.
- [52] Tolhurst G., Heffron H., Lam Y. S., et al. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the g-protein-coupled receptor FFAR2 [J]. *Diabetes*, 2012, 61(2): 364-371.
- [53] Freeland K. R., Wolever T. M. Acute effects of intravenous and rectal acetate on glucagon-like peptide-1, peptide YY, ghrelin, adiponectin and tumour necrosis factor-alpha [J]. *Br. J. Nutr.*, 2010, 103(3): 460-466.
- [54] Samuel B. S., Shaito A., Motoike T., et al. Effects of the gut microbiota on host adiposity are modulated by the short-chain fatty-acid binding G protein-coupled receptor, Gpr41 [J]. *P. Natl. Acad. Sci. USA*, 2008, 105(43):16767-16772.
- [55] Estaki M., Pither J., Baumeister P., et al. Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions [J]. *Microbiome*,



- 2016, 4(1):42.
- [56] Zheng G. H., Li S. Z., Huang M. M., et al. The effect of Tai Chi training on cardiorespiratory fitness in healthy adults: A systematic review and meta-analysis [J]. *PLoS One*, 2015, 10(2):e0117360.
- [57] Matsumoto M., Inoue R., Tsukahara T., et al. Voluntary running exercise alters microbiota composition and increases n-butyrate concentration in the rat cecum [J]. *Bio-sci Biotech Bioch*, 2008, 72(2): 572-576.
- [58] Hoffman-Goetz L., Spagnuolo P. A., Guan J. Repeated exercise in mice alters expression of IL-10 and TNF-alpha in intestinal lymphocytes [J]. *Brain Behav. Immun.*, 2008, 22(2): 195-199.
- [59] Luo B. B., Xiang D., Nieman D. C., et al. The effects of moderate exercise on chronic stress-induced intestinal barrier dysfunction and antimicrobial defense [J]. *Brain Behav. Immun.*, 2014, 39: 99-106.
- [60] Islam H., Townsend L. K., Mckie G. L., et al. Potential involvement of lactate and interleukin-6 in the appetite-regulatory hormonal response to an acute exercise bout [J]. *J. Appl. Physiol.*, 2017, 123(3): 614-623.
- [61] Kasubuchi M., Hasegawa S., Hiramatsu T., et al. Dietary Gut Microbial Metabolites, Short-chain Fatty Acids, and Host Metabolic Regulation [J]. *Nutrients*, 2015, 7(4): 2839-2849.
- [62] Fukiya S., Arata M., Kawashima H., et al. Conversion of cholic acid and chenodeoxycholic acid into their 7-oxo derivatives by *Bacteroides intestinalis* AM-1 isolated from human feces [J]. *Fems. Microbiol. Lett.*, 2009, 293(2): 263-270.
- [63] Porras D., Carbajo-Pescador S., Juarez-Fernandez M., et al. Exercise modulates gut microbiota and intestinal barrier functionality counteracting early obesity and NAFLD in an in vivo model [J]. *J. Hepatol.*, 2019, 70(1): 515-516.
- [64] Meissner M., Lombardo E., Havinga R., et al. Voluntary wheel running increases bile acid as well as cholesterol excretion and decreases atherosclerosis in hypercholesterolemic mice [J]. *Atherosclerosis*, 2011, 218(2): 323-329.
- [65] Hagio M., Matsumoto M., Yajima T., et al. Voluntary wheel running exercise and dietary lactose concomitantly reduce proportion of secondary bile acids in rat feces [J]. *J. Appl. Physiol.*, 2010, 109(3): 663-638.
- [66] Psichas A., Sleeth M. L., Murphy K. G., et al. The short chain fatty acid propionate stimulates GLP-1 and PYY secretion via free fatty acid receptor 2 in rodents [J]. *Int. J. Obesity*, 2015, 39(3): 424-429.
- [67] Doyle A., Zhang G. H., Fattah E. A. A., et al. Toll-like receptor 4 mediates lipopolysaccharide-induced muscle catabolism via coordinate activation of ubiquitin-proteasome and autophagy-lysosome pathways [J]. *Faseb. J.*, 2011, 25(1): 99-110.
- [68] Fischer C. P. Interleukin-6 in acute exercise and training: what is the biological relevance? [J]. *Exerc. Immunol Rev.*, 2006, 12(6):33.
- [69] Starkie R., Ostrowski S. R., Jauffred S., et al. Exercise and IL-6 infusion inhibit endotoxin-induced TNF-alpha production in humans [J]. *Faseb. J.*, 2003, 17(3): 884.
- [70] Munoz-Canoves P., Scheele C., Pedersen B. K., et al. Interleukin-6 myokine signaling in skeletal muscle: A double-edged sword? [J]. *Febs. J.*, 2013, 280(17): 4131-4148.
- [71] Pedersen B. K.. Muscular interleukin-6 and its role as an energy sensor [J]. *Med. Sci. Sport Exer.*, 2012, 44(3): 392-396.
- [72] Furusawa Y., Obata Y., Fukuda S., et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells [J]. *Nature*, 2013, 504(7480): 446.
- [73] Vilorio M., Lara-Padilla E., Campos-Rodriguez R., et al. Effect of moderate exercise on IgA levels and lymphocyte count in mouse intestine [J]. *Immunol Invest.*, 2011, 40(6): 640-656.
- [74] Chambers E. S., Viardot A., Psichas A., et al. Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults [J]. *Gut.*, 2015, 64(11): 1744-1754.
- [75] Sawicki C. M., Livingston K. A., Obin M., et al. Dietary fiber and the human gut microbiota: Application of evidence mapping methodology [J]. *Nutrients*, 2017, 9(2): 125.
- [76] Lira F. S., Rosa J. C., Pimentel G. D., et al. Endotoxin levels correlate positively with a sedentary lifestyle and negatively with highly trained subjects [J]. *Lipids Health Dis*, 2010, 9(1):82.
- [77] Fehrenbach E., Niess A. M., Schlotz E., et al. Transcriptional and translational regulation of heat shock proteins in leukocytes of endurance runners [J]. *J. Appl. Physiol.*, 2000, 89(2): 704-710.
- [78] Dokladny K., Moseley P. L., Ma T. Y. Physiologically relevant increase in temperature causes an increase in intestinal epithelial tight junction permeability [J]. *Am. J. Physiol-Gastr L.*, 2006, 290(2): 204-212.
- [79] Mailing L. J., Allen J. M., Buford T. W., et al. Exercise and the gut microbiome: A review of the evidence, potential mechanisms, and implications for human health [J]. *Exerc. Sport Sci. Rev.*, 2019, 47(2): 75-85.

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